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**TITLE: ANALYSIS OF DENGUE VIRUS ENHANCING EPITOPEs USING
PEPTIDE ANTIGENS DERIVED FROM THE ENVELOPE
GLYCOPROTEIN GENE SEQUENCE**

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FOREWORD

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INTRODUCTION

Increased virus replication in dengue infections can lead to the development of severe disease manifestations of vascular permeability and shock (dengue hemorrhagic fever/dengue shock syndrome, DHF/DSS). Different hypotheses have been proposed as the pathogenic mechanism in DHF/DSS. In the same outbreak, DHF/DSS cases have been associated with both primary and secondary infections; thus implicating the infecting virus strain as being the cause of serious disease (6). Alternatively, more DHF/DSS cases are documented in patients with secondary DEN infections suggesting that pre-existing DEN antibodies exacerbate severe disease outcome by mediating the enhanced replication of the second DEN virus (9). Unidentified host genetic differences are also likely to be associated with development of DHF/DSS cases, since not all primary or secondary DEN infections result in severe disease.

Development of standardized antibody-dependent enhancement assay. Antibody-dependent enhancement (ADE) assay is an *in vitro* system in which DEN virus infection of mononuclear phagocytes in the presence of DEN antibodies is used as laboratory correlates of the sequential infection hypothesis (9). This study uses the principle of ADE to examine the roles of antibody, virus and host cells in DEN replication. In this past year we have developed a standardized ADE assay that permits us to examine the effect of not only antibody on virus replication but, by controlling other variables, examine also the differences in virus strain replication and host susceptibilities.

Rationale for studying pathogenesis of DEN using Caribbean DEN isolates. The development of DHF/DSS is probably not exclusively associated with a single suggested mechanism, but rather is an interplay of several pathogenic factors. Unlike DEN-endemic regions in Southeast Asia, the introduction of DEN-1 CV1636/77 and DEN-2 Jamaica genotypes into the Caribbean region is clearly defined (3,11). In addition, both virus genotypes are temporally associated with the severe DHF/DSS outbreak in Cuba where retrospective epidemiology suggest that DEN-2 DHF/DSS cases were associated with previous exposure to DEN-1 (7).

The nucleotide sequence and deduced amino acid sequence of DEN-2 Jamaica has been completed (4,5). The genomic sequences encoding DEN-1 CV1636/77 proteins is nearly completed (3; Tables 1, 2, 3, and 4). Synthetic peptides consisting of DEN-2 Jamaica E-glycoprotein sequences elicit antibodies that react by ELISA and plaque reduction neutralization (PRNT) to native virus (19).

Chimeric hybrids of DEN-2 E-glycoprotein gene regions have also been constructed and shown to be immunogenic (Chang, unpublished observations). Based on these results, immune sera against synthetic DEN-1 CV1636/77 genomic regions would provide reagents to study if DEN-2 Jamaica replication could be enhanced and thus provide an in vitro explanation for the severe DHF/DSS outbreak as suggested by the epidemiological reports on the Cuba outbreak (7).

ACCOMPLISHEMENTS FOR FY 1990

Selection of a human mononuclear phagocytic cell line for use in ADE assays. Human and monkey peripheral blood leukocytes (PBL) as well as several mononuclear cell lines have been used in ADE assays (2,9,17). We compared the susceptibility of human mononuclear cells to DEN infections using a newly described human promonocytic cell line HL-CZ cells (14, Table 1), K562 (13), U937 (2) cells and cultured human PBLs (Figure 1). DEN-2 16681 virus and 4G2 antibody were added to 3×10^5 of U937, K562 or HL-CZ cells and to 1.5×10^6 HuPBLs. The cultures were incubated for 4 days and harvested by freezing the samples. The amount of productive virus yield was titrated in the BHK-21 clone 15 cell assay (16). The results in Figure 1 represent the amount of virus yield of samples with antibody - samples without antibody (in thousands of pfu/ml). Both HuPBL and U937 could not be infected with less than MOI of 0.01, comparable to results already published elsewhere (2,17). Infection of K562 and HL-CZ cells at MOI of 0.01 however, resulted in high background of virus growth ($>10^3$ pfu/ml) thereby masking the level of enhancement. When the MOI's are lowered to 10^{-4} - 10^{-5} , background virus levels then do not interfere with enhanced virus replication. Thus compared, HL-CZ cell line is more sensitive to DEN virus replication than U937, K562 or HuPBLs.

Characterization of HL-CZ cells. The underlying presumption of ADE is that the presence of antibody mediates increased virus replication by bringing infectious virus closer to the cell surface via the Fc-receptor (FcR). Because of the differences we observed in comparing virus replication with different cells, we hypothesized that there may be a difference in the number of FcRs expressed on these cell surfaces. A direct visual method was developed using sheep red blood cells (SRBC) (BBL, Becton-Dickinson, Cockeysville, MD) sensitized with rabbit anti-SRBC (sSRBC) (BBL, Becton-Dikinson) to enumerate the number cells that express FcRs on their surfaces. A cell that adsorbs three or more sSRBC after a fixed incubation time was counted as a FcR-bearing cell (rosette), the results are listed in Table 2. The number of FcRs expressed by cultured HuPBL after three days' incubation is 60% of a mixed leukocyte population (T cells, B cells, monocytes, and macrophages). Our assumption was that cloned human cell lines would likely have more uniform expression of FcRs; however, only 50% of U937 cells formed rosettes while K562 and HL-CZ cells expressed $>80\%$ rosetting capability. The amount of detectable

virus in ADE cultures appears to be associated with the number of FcRs on the cell surfaces and this may likely determine whether a cell may or may not participate in ADE.

There are three types of human immunoglobulin G FcRs: FcRI, FcRII, and FcRIII each identified by monoclonal antibodies 32.2, IV.3, and 3G8 respectively (20). Our next assumption was that there may be qualitative differences in the types of FcRs expressed on each of the cells that participate in ADE. In order to examine those expressed by HL-CZ cells, we pre-blocked cells with the appropriate FcR type-specific monoclonal antibody (Medarex Inc., West Lebanon, NH) then added ssRBC to the blocked cells to determine the FcR-type (Table 3). HL-CZ appears to have all three FcR types with FcRII > FcRI > FcRIII in contrast to K562 cells that have only FcRII expressed on their surfaces. These observations were then confirmed by flow cytometry analysis using FITC-tagged sheep anti-mouse Fab' (Jackson Immunoresearch Labs, West Grove, PA) instead of ssRBC (Figure 2). The results of the forward angle light scatter (FALS) examination of the FITC-stained cells are presented as comparative histograms of equal cell numbers (top left of each panel) and of equal intensity (FALS 1024, top right) between cells pre-incubated with FcR MAbs (open areas) and cells stained with anti-mouse-FITC (closed areas). K562 cells stained with MAb IV.2 (FcRII) is shown as the control (panel A), K562 cells stained with other FcR MAbs were like the control cells (data not shown). HL-CZ cells stained with all three FcR MAbs to varying degrees with FcRII > FcRIII > FcRI, the latter two are different from our rosette-inhibition studies and will need to be resolved by further analyses.

ADE in HL-CZ cells after FcR blocking. We next asked, if by blocking FcR on HL-CZ cells, will we be able to abrogate virus growth? HL-CZ cells were pre-incubated with either one, three, or no (No Ab and 3H5) specific FcR MAbs. The percentage of available FcR remaining on the cell surfaces were enumerated by rosetting (open circle line, Figure 3). The cells were then infected with DEN-2 virus with the addition of 100 ng of 3H5, and the cultures were harvested on days 2 (open bars) and 4 (shaded bars). Background virus growth (without pre-blocking and without 3H5) was significantly lower than that of the positive control samples (no pre-blocking and with 3H5). Cells that were pre-incubated with FcRI MAb (32.2) resulted in less virus growth than the positive control. Where FcRII, FcRIII or all three FcRs are blocked, the enhancing effect of 3H5 was abrogated. These results correlated with the absence of available FcR sites as determined by rosetting, and suggest that selected FcR types may be involved in mediating enhancement.

Antibody preparation for use in a standardized ADE. Three monoclonal antibodies (MAbs) were selected as the antibody standards. These were originally prepared and characterized by the Walter Reed Army Institute of Research (WRAIR) (10). 4G2 is an IgG_{2a} globulin that is flavivirus- and E glycoprotein-specific

reactive. 3H5 is of IgG₁ subclass that reacts with E glycoprotein as well but is specific for DEN-2. 15F3 is directed against the NS1 protein of DEN-1 belonging to IgG_{2a} subclass. Hybridoma culture fluids from these monoclonals were concentrated by 50% ammonium sulfate precipitation, and purified by Protein A column chromatography. Each Ig fraction was standardized by spectrophotometric protein assay (BioRad, Richmond, CA) to 1 mg/ml and tested for virus reactivity by IFA, ELISA and PRNT.

The specificity of the prepared antibodies were tested in ADE: 4G2, and 3H5 participate in ADE at various concentrations, 15F3 does not mediate ADE of DEN-2 viruses or DEN-1 from Thailand (see following section) and marginally enhances DEN-1 CV1636/77. In previous ADE studies involving DEN infections, it has been pointed out that only antibodies with DEN/flavivirus specificities are involved (2). We confirmed this by substituting matched subclass mouse immunoglobulins derived from mineral oil plastocytomas (MOPC 21, UPC 10; Jackson Lab) instead of 4G2 and 3H5 (Figure 4). The lowest limit of our BHK-21 plaque assay is 7 pfu/ml. The addition of 0.001 ug-10 ug/ml of MOPC 21 or UPC 10 did not enhance DEN-2 replication. Both purified 4G2 and 3H5 mediated enhanced virus production at 100-1000 times higher than background. We also observed that the addition of 1.0-10.0 ug/ml of 4G2 induces enhanced virus replication whereas less amounts of 3H5 (100ng-lug/ml) was needed to mediate ADE. 15F3, though not shown in Figure 4, did not mediate enhanced virus growth.

Selection of DEN viruses as control viruses in ADE. Four virus strains of epidemiological importance were selected to be the control virus strains in our standard ADE (Table 4). The two Thailand strains selected are from endemic DEN regions and are the parental viruses from which attenuated candidates are being tested in vaccine trials (1). In addition, DEN-2 16681 virus is the "prototypic ADE virus strain" since many of the enhancement studies have included this strain (9,17). The Jamaican isolates have been discussed earlier and serve to represent strains from an epidemic region in contrast to the Thailand viruses.

Using a standarized input of virus (MOI = 0.0001; Figure 5) and varying concentrations of purified MAbs, HL-CZ cells were infected, cultured for 4 days, and resulting productive virus assayed. The addition of 4G2 resulted in enhanced replication in every case while 15F3 had minimal effect on DEN-1 CV1636/77 virus replication (>10 fold over background). Regardless of how MAbs affected virus growth, the background growth of each virus and the amount of enhanced growth varied. Jamaican virus strains had a higher background growth than the Thailand viruses, and the enhancement profile of each of the viruses differed. The virus growth yields indicate that each of the DEN viruses, under the same growth conditions, have an intrinsic difference in their ability to replicate in this system. We also noted during these studies that it was important to control the variables carefully if we were to use this system to compare viruses. By varying only the input virus, the enhancement profile changes for DEN-2 Jamaica virus

(Fig. 6). If however at a particular MOI examined that the background virus is already at the threshold level (7 pfu/ml), the enhancement profile does not change, and further diminishing of input virus from that point only results in undetectable virus replication.

Anti-peptide sera directed against DEN-2 E glycoprotein regions. The immune serum obtained from mice immunized with a series of DEN-2 Jamaica synthetic peptides were examined in the ADE system. The DEN-2 synthetic peptide antigens used to immunize animals represent continuous and discontinuous E glycoprotein regions (19; Table 5). Mouse anti-peptide sera were diluted from 10^3 to 10^5 and added to 3×10^5 HL-CZ cells with an input of 0.00001 DEN-2 Jamaica virus. These experiments were done in triplicate and repeated three times, the BHK-21 plaque assay results reported as pfu/ml (Table 5). This data suggest that peptides 1-2 and 437 may be involved in eliciting antibody that mediate enhancement. These observations are preliminary and need to be repeated using purified and standardized quantities of anti-peptide sera.

Cloning and sequencing of DEN-1 CV1636/77. The genome encoding the structural genes of DEN-1 CV1636/77 have been previously published (3). To examine if regions other than those representing the E glycoprotein would be important in DEN pathogenesis, we have proceeded to obtain the sequence of the genome coding for the non-structural proteins of DEN-1 CV1636/77. Using the same cloning methods for obtaining structural region clones, cDNA clones that encompass the nucleotide sequence for the non-structural genome were generated (Figure 7). From the many clones generated, 5 overlapping cDNA clones were selected for sequencing (Figure 7, closed arrows); the sequences of portions of one clone and another short clone (Figure 7, open arrow regions) are being determined.

The nucleotide sequences encompassing NS4a, NS4b, and NS5 genomic regions of DEN-1 are presented along with the published sequences of DEN-2 (5), DEN-3 (18), and DEN-4 (15) in Tables 6, 7, and 8 respectively. The nucleotide region for NS1, NS2a, NS2b, NS3 and the final 800 base pair sequence of the 3'-end of the viral RNA has not been finalized. In Figure 8, a diagrammatic representation of the protein similarities between NS4a, NS4b, and NS5 regions of the DEN viruses are presented (narrow bars represent a single amino acid change, whereas wider bars represent from 2 or more to clusters of amino acid changes). In NS4b, DEN-1 and DEN-4 each have 3 amino acid deletions at amino acid positions 21-23 in comparison with DEN-2 and DEN-3 sequences. Additionally, both DEN-1 and DEN-3 have a deletion at amino acid position 176 of the NS5 protein, the deletion at this position has been verified by RNA sequencing of other DEN-1 and DEN-3 viruses as well (Table 9).

The similarity of the each of the gene regions between the 4 DEN viruses are summarized in Table 10. As expected, DEN-1, DEN-2, and DEN-3 share between 63%-82% similarity over each of the non-structural gene regions. DEN-1 and DEN-4 however are very similar in NS3, NS4a, and NS4b regions (95%-98%); this is contrasted by a

comparison of DEN-1 and DEN-4 over the NS5 region where similarity extends to only 78%. When the combined nucleotide and deduced amino acid sequences over NS3, NS4a, NS4b, and NS5 regions are examined, the similarity of 90% remains between DEN-1 and DEN-4 (Table 11).

CONCLUSIONS

Pathogenic mechanisms by which dengue infections result in serious hemorrhagic manifestations must involve host susceptibility and the infecting virus strain. An animal model in which DHF/DSS can be easily studied does not exist. Therefore in vitro studies of DHF/DSS correlates have primarily depended on ADE experiments. Enhanced replication of virus remains a consistent finding in experimental situations and perhapsr reflect in vivo observations where patients developing DHF/DSS are highly viremic (8).

Relevance of completed research. The development of a standardized ADE assay provides an useful tool to examine the variables that are involved in enhanced virus replication in vitro. We have been able to determine that antibodies of different DEN specificities vary in their ability to mediate enhanced virus growth. We have also determined that viruses have intrinsic differences in their ability to replicate. Our comparison of ADE in different human cell cultures have led to the identification of a cell line that is more analogous to HuPBL in their expresssion of all three FcRs. By using these HL-CZ cells, we have been able to determine that ADE requires the expression of FcRII > FcRIII and probably not as likely to involve FcRI.

Using anti-peptide sera directed against selected DEN-2 E glycoprotein regions, we have been able to demonstrate that some of the E regions will elicit antibody that will mediate enhanced virus replication, neutralization and enhancement, and neutralization alone. Though this series of experiments need to be repeated with purified antibodies, this is the first direct association of specific E-glycoprotein regions with biological functions.

Analyses of the genetic relatedness of DEN viruses. The close relatedness of DEN-1 and DEN-4 in the NS3-NS4a,b regions; the shared deletions of DEN-1 and DEN-3 in NS5 identifies specific genomic regions of interest to study in relating genomic sequence similarity/differences relate to virus replication and antigenicity.

The comparison of the nucleotide sequences with the other DEN virus strains have been very interesting. It was expected that the similarities between DEN serotypes would extend between 60-80%. The high percentage of shared sequences in NS3, NS4a, NS4b between DEN-1 and DEN-4 suggests two scenarios. The first explaination may be that genomic recombination has occured and the second more likely explaination may be that DEN-1 and DEN-4 have evolved from a common progenitor. Why these relationships have not been

detected before is not surprising since the serological tests commonly used to differentiate the viruses are directed toward the structural proteins; alternatively, because the non-structural regions are not expressed, there is little selective pressure to restrain these genomic regions.

OBJECTIVES FOR FY 1991

1. Completion of the sequencing and analyses of DEN-1. The NS1, NS2a, NS2b, and the 3'-end of the DEN-1 sequence will be completed. We will compare the nucleotide sequence and the deduced amino acid sequence with the other DEN serotypes.
2. Identify the genomic regions that elicit antibodies that elicit neutralization/enhancement. Extend our preliminary findings using purified antibodies. Elicit antibodies to new DEN-1 and DEN-2 synthetic antigens, purify the antibodies and examine their reactivities in PRNT and ADE.
3. Examine genomic variation of DEN strains. Determine if DEN virus strains share similar nucleotide sequence over the genomic regions that mediate neutralization/enhancement. We will do primer-directed RNA sequencing of selected DEN strains that are associated with DF or DHF/DSS patients.
4. Confirm the relevance of defined neutralization/enhancement epitopes to human infections. Patient serums will be used to react with the synthetic antigens that define neutralization and/or enhancement activity.

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TABLE 1. HUMAN PROMONOCYTIC CELL LINE
HL-CZ CLONE CCC-5

- 50 YR. OLD MALE WITH ADULT T-CELL LEUKEMIA
- PROMONOCYTIC CELL LINE THAT SHOW EARLY DIFFERENTIATION IN HEMATOPOIESIS
- PHAGOCYTIC, WITH SINGLE, DOUBLE, OR MULTIPLE NUCLEI
- 90% OF THE CELLS POSSESS CD-15 MARKER

- LIU, W.T. et al. (1989) J BIOL MED SCI
4:284

Table 2. DETECTION OF Fc RECEPTORS (FcR) OF HUMAN MONONUCLEAR CELLS

CELLS	KNOWN FcR TYPE (a)	% ROSETTES (b)
HUMAN PBL	I, II, III	60
U937	I, II	50
K562	II	87
HL-CZ	??	87

(a) Unkeless et al., Ann Rev Immunol 6(1988):251-81.

(b) Sheep red blood cells (SRBC) sensitized with rabbit anti-SRBC (sSRBC). 10^4 human mononuclear cells were incubated with sSRBC at 4C for 12 hours. Percent rosettes are determined by the number of cells with sSRBC/total number of mononuclear cells counted. Total number of mononuclear cells counted = 100 to 400 cells.

TABLE 3. Fc RECEPTOR (FcR) DETECTION BY
ROSETTE INHIBITION

<u>Fc CLASS</u>	<u>K562</u>	<u>HL-CZ</u>
FcRI (MAb 32.2)	165/197 (84%)	24/178 (14%)
FcRII (MAb IV.3)	0/165 (0)	1/142 (1%)
FcRIII(MAb 3G8)	186/278 (70%)	70/182 (38%)
NO BLOCKING AB	141/162 (87%)	183/209 (86%)

TABLE 4.

ISOLATES FROM DENGUE
PATIENTS

STRAIN I.D.	YEAR
DEN- 1 THAILAND 16007	DF 1964 (1)
DEN- 2 THAILAND 16681	DHF 1964 (1)
DEN- 1 JAMAICA CV1636/77	DF 1977 (2)
DEN- 2 JAMAICA 1409	DF 1983 (3)

(1) Halstead et al. 1970. Yale J Biol Med 42:2(2) King et al. 1979. PAHO 375:153.(3) Deubel et al. 1986. Virol 165:234.

Table 5. ACTIVITY OF MOUSE ANTI-DENGUE 2 PEPTIDE IN NEUTRALIZATION AND ADE ASSAYS

PEPTIDE I.D.	COMPRISED OF E AMINO ACID #	PEPTIDE LENGTH	PRNT (a) TITER	ADE (b) TITER
1-2	1-30	30	160	132
35	35-55	22	20	< 7
3-8/2	49-60, 121/140	32	160	< 7
3-8/1	58/73, 106-115, 117/121	31	40	< 7
4-6	72-91, 93-105	33	40	13
79	79-99	21	40	< 7
5-7	90-104, 106-120	30	160	< 7
04	121-140	20	40	< 7
142	142-172	32	40	< 7
142-1	165-172	9	40	< 7
208	208-219	13	160	< 7
06	225-249	26	20	< 7
67	255-274	21	160	< 7
07	302-333	32	20	< 7
16	333-351	22	80	< 7
17	352-368	18	20	< 7
388	388-400	14	20	< 7
437	437-452	17	40	200
CONTROL	3H5 10 NG/ML	NA (c)	--	1466
CONTROL	4G2 100 NG/ML	NA	--	400

(a) PRNT, 70% plaque reduction.

(b) ADE, 0.00001 MOI, HL-CZ cells. Average of triplicate samples expressed as pfu/ml. Negative control background = < 7 pfu/ml.

(c) NA, not applicable.

TABLE 6. NUCLEOTIDE SEQUENCE OF DEN-1 CV1636/77
Nucleotide sequence encoding NS4a genome

TABLE 6. NS4a nucleotide sequence (Page 2)

DEN-1	AGG ACC CCA CAA GAC AAT CAA TTG ATC TAC GTC ATA TTG ACC	420
DEN-2	..A ..A ..CCC.T G.C A.A G..	
DEN-3	..A ..T ..CC ... C.C GCA ..T ... G.G A.A GG.	
DEN-4	462
DEN-1	ATT CTC ACC ATC ATT GGT CTA ATA GCA GCC	
DEN-2	..CA G.G G.G .CC GC. .CC ATG ..A	
DEN-3	..A ..T ..A T.G GC. .CA A.. G.. .G ...	
DEN-4	

TABLE 7. NUCLEOTIDE SEQUENCE OF DEN-1 CV1636/77
Nucleotide sequence encoding the NS4b genome

DEN-1	ACC GAG ATG GGG CTG ATT GAA AAA ACA AAA ACG GAT TTT GGG	42
DEN-2	.A.T T.C C.GC ..G .AA ..C C.C ..A	
DEN-3	.AT ..AA ... T.GCTG .GAA ..A	
DEN-4	.A.AAAAAAA	
		84
DEN-1	TTT TAC CAG GTA AAA ACA	GAA ACC ACC ATC CTC
DEN-2	..G GGA AGC A.T .C. ..T CAG GAA TCT	..G .G. .A.G
DEN-3	A.G .CT A.A .A. CC. GGT GTT GTT TCT CC.G. TAT T.G
DEN-4A.AAAAAA	
		126
DEN-1	GAT GTG GAC TTG AGA CCA GCT TCA GCA TGG ACG CTC TAT GCA	
DEN-2	..C A.A ..T C.A CGT ..T ..AAAGC
DEN-3A.AAAAAA	T.G ..C ..C
DEN-4A.AAAAAA	
		168
DEN-1	GTA GCC ACC ACA ATT CTG ACT CCC ATG CTG AGA CAC ACC ATA	
DEN-2	..G ..T ..A ... T.. G.T ..A ..A ... T..T .G. ..T	
DEN-3	..GA ... G.A A.A ..A ..A ... T..A	
DEN-4A.AAAAAA	
		210
DEN-1	GAA AAC ACG TCG GCC AAC CTA TCT CTA GCA GCC ATT GCC AAC	
DEN-2T T.C ..A .TG ..T G.G ..C ... A..AT	
DEN-3	..G ..T T.C A.A ..A ..T G.G ..C ..G ..AA ..T	
DEN-4A.AAAAAA	
		252
DEN-1	CAG GCA GCC GTC CTA ATG GGG CTT GGA AAA GGA TGG CCG CTC	
DEN-2	..A ..C A.A ..GATGAA T.G	
DEN-3TGGT T.A .ACAA	
DEN-4A.AAAAAA	
		294
DEN-1	CAC AGA ATG GAC CTC GGT GTG CCG CTG TTA GCA ATG GGA TGC	
DEN-2	TCA .AG ..C C.. A.. ..A ..T ..C ..C C.T ..C ..T	
DEN-3	TCG .A.T.G ..C ..A ..A ..A ..G ... C.. ..T	
DEN-4A.AAAAAA	
		336
DEN-1	TAT TCT CAA GTG AAC CCA ACA ACC TTG ACA GCA TCC TTA GGC	
DEN-2	..C ..A ..G ..CT .T. ... T C.CG.T C.T CTT	
DEN-3A	
DEN-4A.AAAAAA	
		378
DEN-1	ATG CTT TTA GTC CAT TAT GCA ATA ATA GGC CCA GGA TTG CAG	
DEN-2	T.A T.G G.. .CAAC ..TGA C.T ..A	
DEN-3	T.. ..A G.C ACAATTT	
DEN-4A.AAAAAA	
		420
DEN-1	GCA AAA GCC ACA AGA GAG GCC CAG AAA AGG ACA GCT GCT GGG	
DEN-2A.AAAAAAA	
DEN-3A.AAAAAA	
DEN-4A.AAAAAA	

TABLE 7. Nucleotide sequence of NS4b (Page 2)

DEN-1	ATC ATG AAA AAT CCC ACA GTG GAC GGG ATA ACA GTA ATA GAT	462
DEN-2C ..AC ..T ..AG ..T ..C	
DEN-3	..AGA ..GT ..ATG AC.C	
DEN-4	
		504
DEN-1	CTA GAA CCA ATA TCC TAT GAC CCA AAA TTT GAA AAG CAA TTA	
DEN-2T C..TG ...	
DEN-3T ..T G.. ATAT T.. C..	
DEN-4	
		546
DEN-1	GGG CAG GTC ATG CTA CTA GTC TTG TGT GCT GGA CAA CTA CTC	
DEN-2	..A ..A ..AC ... A.. C.C ..C .TG ACT ... G.. T.A	
DEN-3	..A ..G ...TC ..G ...T C..A .TTT T.G	
DEN-4	
		588
DEN-1	TTG ATG AGA ACA ACA TGG GCT TTC TGT GAA GTC TTG ACT TTG	
DEN-2	A..G ..T C.GG .CT C.A ..C ..A	
DEN-3	..A T..C ..GT CAT ..C C.A	
DEN-4	
		630
DEN-1	GCC ACA GGA CCA ATC TTG ACC TTG TGG GAG GGC AAC CCG GGA	
DEN-2	..G ..C ..G ..TCC ..A C..A ..A ..T ..A ..G	
DEN-3A ACA ..A C.CAA ..A TCA ..T ..G	
DEN-4	
		672
DEN-1	AGG TTT TGG AAC ACG ACC ATA GCC GTA TCC ACC GCC AAC ATT	
DEN-2TT ..A ..G ..A .TG ..TC	
DEN-3	.A. ..CC ..GT ..TTG ..GC	
DEN-4	
		714
DEN-1	TTC AGG GGA AGT TAC TTG GCG GGA GCT GGA CTG GCT TTT TCA	
DEN-2	..T ..A ..G ..CCT CTCC	
DEN-3A ..G ..C ..T ..A ..AG ..T ... C.. ..T	
DEN-4	
		744
DEN-1	CTC ATA AAG AAT GCA CAA ACC CCT AGG AGG	
DEN-2	A.. ..G C A.. AC. ..A. A.A ..A ..A	
DEN-3	A.. ..G ..A TCA .TT GG. ..A GGA .A. ..A	
DEN-4	

TABLE 8. NUCLEOTIDE SEQUENCE OF DEN-1 CV1636/77
Nucleotide sequence encoding the NS5 genome

DEN-1	GGA ACT GGG ACC ACA GGA GAG ACA CTG GGA GAG AAA TGG AAA	42
DEN-2 C .A. .T. .	
DEN-3 A . . . T.A CA. . . T .A . . C T.A A . . G . . .	
DEN-4	... G . . .	
		84
DEN-1	ACA CAG TTA AAC CAA CTG AGC AAG TCA GAA TTC AAC ACC TAC	
DEN-2	.GC .GA . .G . . . GC. . . . G.A ..A AGT T C.G .T. .G	
DEN-3	.AG A.A T .G T.A TC. CG. AA. . . G . . T G. . CTT . .	
DEN-4	.G. . . C. . . . TC. T.A GA. .GA AA. . . G . . T G.A GAG . . T	
		126
DEN-1	AAA AGG AGT GGG ATT ATG GAG GTG GAC AGA TCC GAA GCC AAA	
DEN-2	..G .AA . . . A . .C CA. . . A G . . . A.. TT. . . A . .	
DEN-3	..G .AA TCC ..A . .C .CC ..A T . . . A.A ..A	
DEN-4A A . .A C.A ..A G A.T G	
		168
DEN-1	GAG GGA TTG AAA AGA GGA GAA ACA ACC AAA CAT GCA GTG TCG	
DEN-2	..A ..C A.C G GA. C.C ..C ..T	
DEN-3	..A ..G ..A T. . . A C.C C C	
DEN-4	TCT .CC C. . . . GAT ..G TCT .A. .TG ..G A . . T	
		210
DEN-1	AGA GGA ACA GCC AAA CTG AGG TGG TTT GTG GAG AGG AAC CTC	
DEN-2	C.. . .C T.. . .A A C ..C A . . T A.G	
DEN-3C .GC ..A T CAA C A . . . A.G	
DEN-4G T.C GCT ..G A.C ..A . . . A.. . . T . . . A GGG A.G	
		252
DEN-1	GTG AAA CCA GAA GGG AAA GTC ATA GAC CTC GGT TGT GGA AGA	
DEN-2	..C .C. G ..G G.G C ..C . . .	
DEN-3	..C .TT ..C A .G. T.A ..C	
DEN-4	..A ..G . . . A.. . . . G.. . . T G.. . . T ..T ..C G	
		294
DEN-1	GGT GGC TGG TCA TAT TAT TGT GCT GGG CTG AAG AAA GTC ACT	
DEN-2	..G C .	
DEN-3	..A C .	
DEN-4	..A ..A T C ATG ..G ACA ..C C ..G . .	
		336
DEN-1	GAA GTG AAG GGA TAC ACA AAA GGA GGA CCT GGA CAT GAG GAA	
DEN-2C ..A ..C CTG .	
DEN-3 CGA G. C ..C ..A C ..A . . .	
DEN-4 A ..G ..T .	
		378
DEN-1	CCT ATC CCA ATG GCG ACC TAT GGA TGG AAC CTA GTA AAG CTA	
DEN-2	..C C . . . T.A ..A G	
DEN-3	..A G.A ..T . . . T.T ..A ..C	
DEN-4	..C ..T ..C T ..T T	
		420
DEN-1	CAC TCT GGA AAA GAT GTA TTT TTC ACA CCA CCT GAG AAA TGT	
DEN-2	..A AG. . . . GTT ..C ..T ..C C ..C ..A ..A ..G . . .	
DEN-3	ATG AG. G . . . C.T AT CTG A ..G . . .	
DEN-4	..T ..A ..G GTT ..C T.G ..C .AC ..A. . . C A.A . . . C.. GTG	

TABLE 8. NS5 nucleotide sequence (Page 2 of 7)

DEN-1	GAT ACC CTT CTG TGT GAT ATT GGT GAG TCC TCT CCG AAT CCA	462
DEN-2A T.G T.. . .C ..A ..GG ..A ..AC	
DEN-3A T.. . .C .. .A ..A ..T ..A ..A ..G C ..	
DEN-4	..C .. .G ..CG .. .A .. T.T	
		504
DEN-1	ACT ATA GAA GAA GGA AGA ACG TTA CGT GTT CTA AAG ATG GTG	
DEN-2	..GC. ... C.. ..A C.C A.A ..C ..C ..C T.A ..A	
DEN-3	..A G.GA.CC A.. A.A ..C T.GT	
DEN-4	..AGA .. A.A ... T.G	
		546
DEN-1	GAA CCA TGG CTC AGA GGA AAC	
DEN-2	... AAT ... T.G .AC AAT ... ACCTG G.T	
DEN-3A .A. AACG ..T ..C ..T ... G.A	
DEN-4	..GTCT TC. ..A CCT G.. . . .G ..C ... G..	
		588
DEN-1	CTA AAT CCT TAC ATG CCA AGT GTG GTA GAA ACT CTG GAG CAA	
DEN-2	..C ..C ..A ..TC TCA ..C A.. . .AA A.. ..A AC.	
DEN-3	TTG ..C ..AC. ... A.T ..G CAC T.A ..A AG.	
DEN-4	..T ..C ..CCA ..C A.. . .GAGA..	
		630
DEN-1	ATG CAA AGA AAA CAT GGA GGG ATG CTA GTG CGA AAC CCA CTC	
DEN-2	C.AG ... T.. . . .A GCC T.. . . .A.G ..T	
DEN-3	C.AAT ... A.. ..T	
DEN-4	C.. .GT ..TAC ..T ..C A.. TG. ..G ..G	
		672
DEN-1	TCA AGA AAT TCC ACC CAT GAA ATG TAC TGG GTT TCA TGT GGA	
DEN-2	... C.. . .CAGA ..C AA. .C.	
DEN-3	... C.. . .CCT ... A.A ..C AA. ..T	
DEN-4	..C ..G ..CGTG ... G.A ..CG	
		714
DEN-1	ACA GGA AAC ATT GTG TCG GCA GTG AAC ATG ACA TCC AGA ATG	
DEN-2	T.C ..GAA T..TT ..A ..G ...	
DEN-3CC ..C ..T T.. . .CGT.T..	
DEN-4	T.GAGC T.TCAA ..AG ...	
		756
DEN-1	TTA CTG AAT CGA TTC ACA ATG GCT CAC AGG AAG CCA ACA TAT	
DEN-2	..G A.T ..C A..AAAA. ..A G.C ..C ..C	
DEN-3	C.. . . .C A..A.AGA ..C ..C ATA	
DEN-4	..G T.. . .C A.GCA AGG ..TA ..C ..T ...	
		798
DEN-1	GAA AGA GAC GTG GAC TTA GGC GCT GGA ACA AGA CAT GTG GCA	
DEN-2	..G .C. ..T ..T ... C.. ..A AGCC C.C A.C A.T ..G.	
DEN-3	..G .A. ..TTA ..AC C.. . .AC AAT	
DEN-4	..G .AGA ..T C.T ..G ..AG ... AG. ..C T.C	
		840
DEN-1	GTG GAA CCA GAG GTA GCC AAC CTA GAT ATC ATT GGC CAG AGG	
DEN-2	A.T ... AGT ... A.. C.A ..TC ..A ..A ..A A.. ..A	
DEN-3	.C.A AC. C.. . .A.G ... G.. . .G ..G G.A ..A	
DEN-4	ACT ... A.. A.A AA. C.A G.. A.G ACAG. ..G AGA ...	

TABLE 3. NS5 nucleotide sequence (Page 3 of 7)

DEN-1	ATA GAG AAT ATA AAA AAT GAA CAC AAG TCA ACA TGG CAT TAT	882
DEN-2 A . . . G C.A . . G . . T G.A A. . T. . . . C . .	
DEN-3	... A.A . GG . . C . . . G.G . . G . . T . GT C . .	
DEN-4	C.T C. . CGA T.G C. . G.A . . G . . . A GA. . . C	
		924
DEN-1	GAT GAA GAC AAT CCA TAC AAA ACA TGG GCC TAT CAT GGA TCA	
DEN-2	.. C C. . . C.C G T . . C . . . C AGC	
DEN-3 T . . A . . . T G T . . C C	
DEN-4	... C.G . . A . . C . . A . . . G. . . C . . . G . . T . . . AGC	
		966
DEN-1	TAT GAG GTC AAG CCA TCA GGA TCA GCC TCA TCT ATG GTG AAT	
DEN-2 A ACA . . A . A.T A C	
DEN-3 A . . A . . A G.C A. . . C C . . C . . . A.A . .	
DEN-4 A CCT CCT T.G A. . . C . . T C.A . . C C	
		1008
DEN-1	GGC GTG GTG AGA TTG CTC ACA AAA CCA TGG GAT GTT ATC CCC	
DEN-2	.. A . . . C . . C. . . G C C . . C G. . . T	
DEN-3	.. A . . C . . . A. . C.C G G.G . . A	
DEN-4	.. G . . A . . A. . C.C . . A C G . . T . . A	
		1050
DEN-1	ATG GTC ACA CAA ATA GCT ATG ACT GAT ACC ACA CCC TTC GGA	
DEN-2 G . . . G . . G . . A A . . C . . G . . T . . A . . T . .	
DEN-3 G . . . G . . G . . A A . . . A . . T . . A . . T . . C	
DEN-4 G . . T . . G T. . . C G. . . A . . . G.A . . T C . . A T . . T . . C	
		1092
DEN-1	CAA CAC AGA GTG TTT AAA GAG AAA GTT GAC ACG CGC ACA CCA	
DEN-2 A C.C . . T . . C G A.A . . C . . A.	
DEN-3	.. G . . A . . . T G C A.G T	
DEN-4 A C G . . G . . T . . C A.A	
		1134
DEN-1	AGA GCA AAA CGA GGC ACA GCA CAA ATT ATG GAG GTG ACA GCC	
DEN-2	GA. C.G . . G GA. AAG A. . C.G . . . A.A A.C . . G . . A	
DEN-3	.. G C.C . . TG . C. . . A . . AG. A.G G. A.C G	
DEN-4	CA. C. . . . CC . . T . . CG. ATG G. . . . ACC AC.	
		1176
DEN-1	AAG TGG TTA TGG GGT TTC CTT TCC AGA AAC AAA AAA CCC AGA	
DEN-2	G. . . . C.T . . . AAA GAA . . A GGA . . AG . . A . . G . . C. . . T . . G	
DEN-3	G. . . . C.T . . . A.G AC. . . G GGA . . G G.	
DEN-4	.. T . . C.G . . . CC C. . . . GGA . . AG . . G T	
		1218
DEN-1	ATC TGC ACA AGA GAG GAG TTC ACA AGA AAG GTT AGG TCA AAC	
DEN-2	.. G . . T . . C . . . A . . A G . . A AGC . . T	
DEN-3	T.A G AG C . . A A.C . . .	
DEN-4	C.G G . . A TC TCA . . A A	
		1260
DEN-1	GCA GCA ATA GGA GCA GTG TTC GTT GAT GAA AAC CAA TGG AAC	
DEN-2 C T.G . . G . . C A.A . . . AC. G . . A A	
DEN-3 T . . G . . C . . T . . C . . . ACA . . A . . G G..	
DEN-4 C . . A . . T CGA . . C . . T CAG . . A . . A C.G GG. . . . CA	

TABLE 3. NS5 nucleotide sequence (Page 4 of 7)

DEN-1	TCA GCA AAA GAA GCA GTG GAA GAC GAA AGG TTT TGG GAT CTC	1302
DEN-2	..G ... CGT ..G ..T ..TT AGTG ..G	
DEN-3	AGT ..G .G. .CT ..T ..T ..G ..C ... GAA A.A ..T	
DEN-4C .GTT ... A.T ... AGC C..A ..G	
		1344
DEN-1	GTG CAC AGA GAG AGG GAC CTT CAT AAA CAG GGA AAA TGT GCC	
DEN-2	..T G.. .G ..A ..A T ..C ... CTT G.AGA	
DEN-3	... G.. . . .A C.T . A ..C ..CT. ..C ..GGA	
DEN-4	..T G.. .A. ..ACC ..A ..C CAG G.A ..GAA	
		1386
DEN-1	ACG TGT GTC TAC AAC ATG ATG GGG AAG AGA GAG AAA AAA TTA	
DEN-2	..AGA ..AG ... C..	
DEN-3	.GC ..C ..TC ..GC.T	
DEN-4	T..TA ..A C.TG ...	
		1428
DEN-1	GGA GAG TTT GGA AAG GCA AAA GGA AGT CGT GCA ATA TGG TAC	
DEN-2	..GC ..CT ..C A.A ..CG	
DEN-3T ..AC ... A.G ..T	
DEN-4C .GA ..C ..GC ..AC ...	
		1470
DEN-1	ATG TGG CTG GGA GCA CGC TTT CTA GAC TTC GAA GCC CTT GGT	
DEN-2TC T..TA ..A	
DEN-3 T.. . . .C A.G ..AC ..TC ..G ..G ..C ..A	
DEN-4G ..GG ..A ..TG ...	
		1512
DEN-1	TTC ATG AAT GAA GAT CAC TGG TTC AGT AGA GAG AAT TCA CTC	
DEN-2	... T.. TCCG ..C ..C ..G	
DEN-3	... C.CC TCG C.T ..A ..C ..T TA.	
DEN-4	... T.. G.CA TGG	
		1554
DEN-1	AGT GGA GTG GAA GGA GAA GGA CTG CAC AAA CTT GGA TAC ATA	
DEN-2GG ..A ..CT	
DEN-3 AAG ..GC	
DEN-4GTG. T.GT ..C	
		1596
DEN-1	CTC AGA GAC ATA TCA AAG ATT CCG GGG GGA AAT ATG TAT GCA	
DEN-2	T.A G.G AGCAG GAA GCAC ..C	
DEN-3	TTGT ..T ..CA ..C ..AGCCT	
DEN-4	..G GAG ..G ... GACAG GAT ..A ..AC CTAT	
		1638
DEN-1	GAT GAT ACA GCC GGA TGG GAC ACA AGA ATA ACA GAG GAT GAT	
DEN-2CAC ... CTA ..A ..C	
DEN-3CTAC	
DEN-4CA ..CC ..TC	
		1680
DEN-1	CTT CAG AAT GAG GCT AAA ATC ACT GAC ATC ATG GAG CCT GAA	
DEN-2	T.A A.AA .AA .TG GTA ..A A.. CA.A CGA ...	
DEN-3	..G ..CAAA C.G CAGC	
DEN-4AAC CTGG ..A CAGCT ..C CAC	

TABLE 8. NS5 nucleotide sequence (Page 5 of 7)

DEN-1	CAT GCT CTA TTG GCT ACG TCA ATT TTT AAG CTG ACC TAC CAA	1722
DEN-2	..C AAG AA. C.A ..C GA. G.C ..A ..C ..A T.A ..G	
DEN-3	..C AGG CAG C.A ..G .AC G.T ..A ..CC ..A	
DEN-4	..C AAG ATC C.A ..C .AA G.CC ..A ..AT ...	
		1764
DEN-1	AAC AAG GTG GTG ACC CTG CAA AGA CCA GCA AAA AAT GGA ACC	
DEN-2 CGT G.. A.. CC. .GA ..C ..A	
DEN-3AC .AA G.C ... C.. ... A.T CC. ..G ..C ..G	
DEN-4A TTT G.C CTCC A.. CCG .GA ... G.G	
		1806
DEN-1	GTG ATG GAT GTT ATA TCC AGA CGT GAC CAG AGA GGA AGT GGA	
DEN-2	..A A.CG ... A.AACG	
DEN-3	..AC A.CT ..G AAAAC	
DEN-4	..A A.CG AAA ..A ..AT	
		1848
DEN-1	CAG GTC GGA ACT TAT GGC TTA AAT ACT TTC ACC AAT ATG GAG	
DEN-2	..AC ..C C.TACA	
DEN-3GT C.G ..C ..ACA	
DEN-4	..A ..TAT ..G ..C ..ACA	
		1890
DEN-1	GTC CAA CTA ATA AGA CAA ATG GAG TCT GAA GGA GTC ATC ACA	
DEN-2	.CT ... T.. ..TG GGA A.. T.. .A.	
DEN-3	.C. ..GCA GGAC ..G T.G T..	
DEN-4	..TC .GC C.CA G..	
		1932
DEN-1	CAA GAT GAC ATG CAG AAC CCA AAA GGT TTG AAA GAA AGA GTT	
DEN-2	AGC AT. C.G CAC .T. .CA GTC .C. .AA GAA .TC .CT GT. CAG	
DEN-3	A.G .CA ... C.C G..C C.T CCG C.A G.G A.G .A. A..	
DEN-4G	
		1974
DEN-1	GAG AAA TGG TCG AAA GAG TGT GGT GTC GAC AGG CTG AAA AGA	
DEN-2	A.CTA GC. AGA GTG ..G CGT ..AA TC. ...	
DEN-3	ACA C..T. G.. ACT AAA ..A ..G ..G ... T.A	
DEN-4 CT. T.A ..G ..G	
		2016
DEN-1	ATG GCA ATT AGC GGA GAT GAT TGT GTG GTG AAA CCA ATT AGT	
DEN-2C ..C ..TT ..AT T.A GA.	
DEN-3C ..CGC ..AC GAC	
DEN-4C ..TCCG ..C C.A GA.	
		2058
DEN-1	GAC AGG TTC GCA ACA GCC TTA ATA GCT CTG AAT GAC ATG GGA	
DEN-2A ..TGT ..TC.A	
DEN-3AC .AT ... CTG C.T ..CC ..T	
DEN-4	..GT .GC ..T T.. C.C C.C TTC T.. ..C C.G	
		2100
DEN-1	AAA GTA AGA AAA GAC ATA CCG CAG TGG GAA CCT TCA AAA GGA	
DEN-2T ..GTAA ..A G. ...	
DEN-3	..G ..T ..G ..GT ..A ... C.G ..AG ...	
DEN-4	..G ..G ..GT A ..T ..G ...	

TABLE 8. NS5 nucleotide sequence (Page 6 of 7)

TABLE 8. NS5 nucleotide sequence (Page 7 of 7)

2562

DEN-1

DEN-2 GGA AAA AGA GAA GAC CAA TGG TGC GGC TCA TTG ATT GGG CTG
 DEN-3 GGA AAG AGA GAA GAC CAA TGG TGC GGA TCA CTC ATA GGT CTC
 DEN-4 GGG AAA AGA GAG GAT TTG TGG TGT GGA TCC CTG ATT GAA CTT

2604

DEN-1

DEN-2 ACA AGC AGG GCT ACC TGG GCA AAG AAC ATC CAA ACA GCA ATA
 DEN-3 ACT TCC AGA GCA ACC TGG GCC CAG AAC ATA CTC ACA GCA ATC
 DEN-4 TCT TCC AGA GCC ACC TGG GCG AAG AAC ATT CAC ACG GCC ATA

2646

DEN-1

DEN-2 AAT CAA GTC AGA TCC CTT ATA GGC AAT GAG GAA TAC ACA GAC
 DEN-3 CAA CAG GTG AGA AGC CTC ATA GGC AAT GAA GAG TTT CTG GAC
 DEN-4 ACC CAG GTC AGG AAC CTG ATC GGA AAA GAG GAA TAC GTG GAT

2688

DEN-1

DEN-2 TAC ATG CCA TCC ATG AAG AGA TTC AGA AGG GAA GAG GAA GAG
 DEN-3 TAC ATG CCT TCG ATG AAG AGA TTC AGG AAG GAG GAG GAG TCA
 DEN-4 TAC ATG CCA GTA ATG AAA AGA TAC AGT GCT CCT TCA GAG AGT

2703

DEN-1

DEN-2 GCA GGT GTC CTG TGG
 DEN-3 GAG GGA GCC ATT TGG
 DEN-4 GAA GGA GTT CTG

TABLE 9. VERIFICATION OF THE DELETION
IN THE NS5 1055-1065 REGION

NUCLEOTIDE SEQUENCES		PRIMER-DIRECTED RNA SEQUENCES	
DEN-1	AGA GGA AAC --- CAA TTC TGC		
DEN-2	AAC AAT AAC AAC CAA TTT TTC		
DEN-3	AAA AAC AAC --- CAG TTT TGC		
DEN-4	TCT TCA AAA CCT GAA TTC TGG		
CV1636	AGA GGA AAC --- CAA UUC UGC		
454-1	AGA GGA AAC --- CAA UUC UGC		
16007	AGA GGA AAC --- CAA UUC UGC		
155-3	AAA AAC AAC --- CAG U		
035-3	AAA AAC AAC --- CAG U		

TABLE 10.

COMPARISON OF DENGUE
NON-STRUCTURAL PROTEINS

	DEN- 2 JAMAICA	DEN- 3 H87	DEN- 4 DOMINICA
DEN- 1			
CV 1636/77			
NS3	63%	72%	95%
NS4a	65%	60%	98%
NS4b	80%	79%	98%
NS5	77%	82%	78%

TABLE 1¹.
**SIMILARITY BETWEEN ALIGNED NUCLEOTIDE
 AND AMINO ACID SEQUENCES OF THE NS3,
 NS4a, NS4b, AND NS5 REGIONS OF DEN**

		NUCLEOTIDE			
		D1	D2	D3	D4
AA		D1	—	70%	69%
	D1	—	—	—	90%
	D2	74%	—	63%	66%
	D3	73%	70%	—	68%
	D4	91%	74%	78%	—

COMPARISON OF ANTIBODY-DEPENDENT
ENHANCEMENT IN DIFFERENT CELLS

MAb 4G2 DEN-2 16681

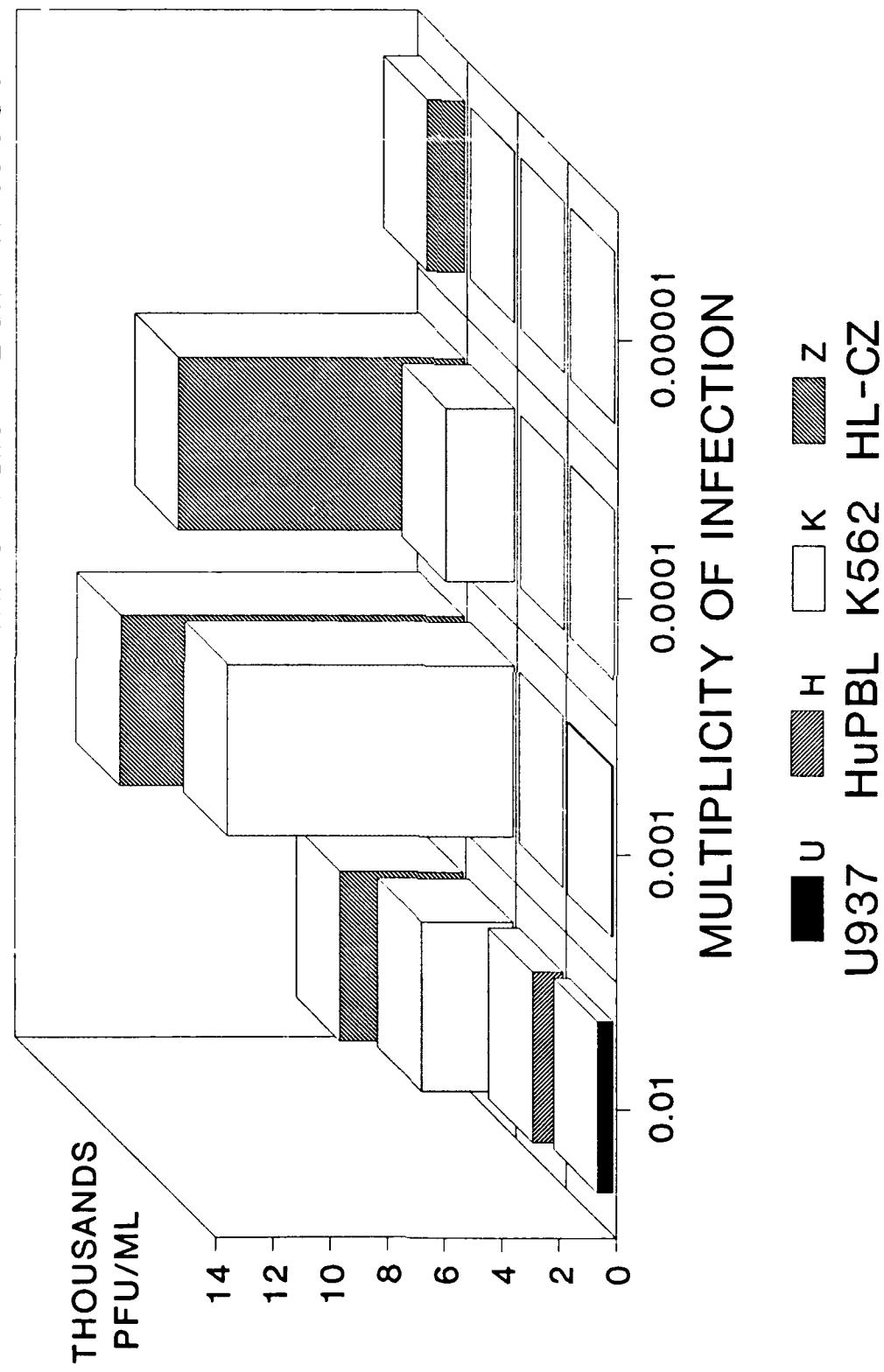


FIGURE 1.

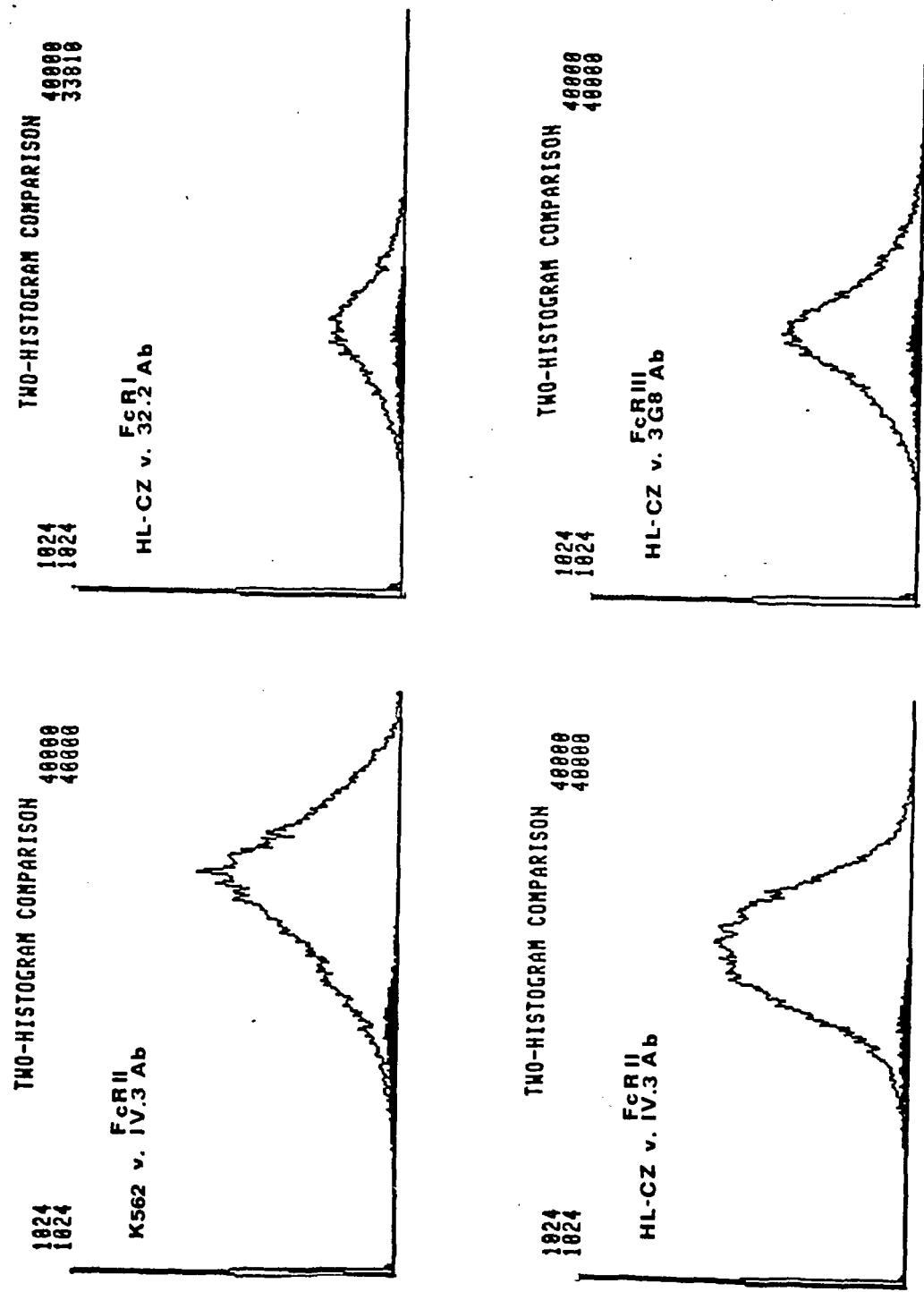


FIGURE 2. ○ CELLS & TEST Ab ● CELLS & CONTROL Ab

EFFECT OF Fc-RECEPTOR MABS IN BLOCKING ADE OF DEN-2 16681

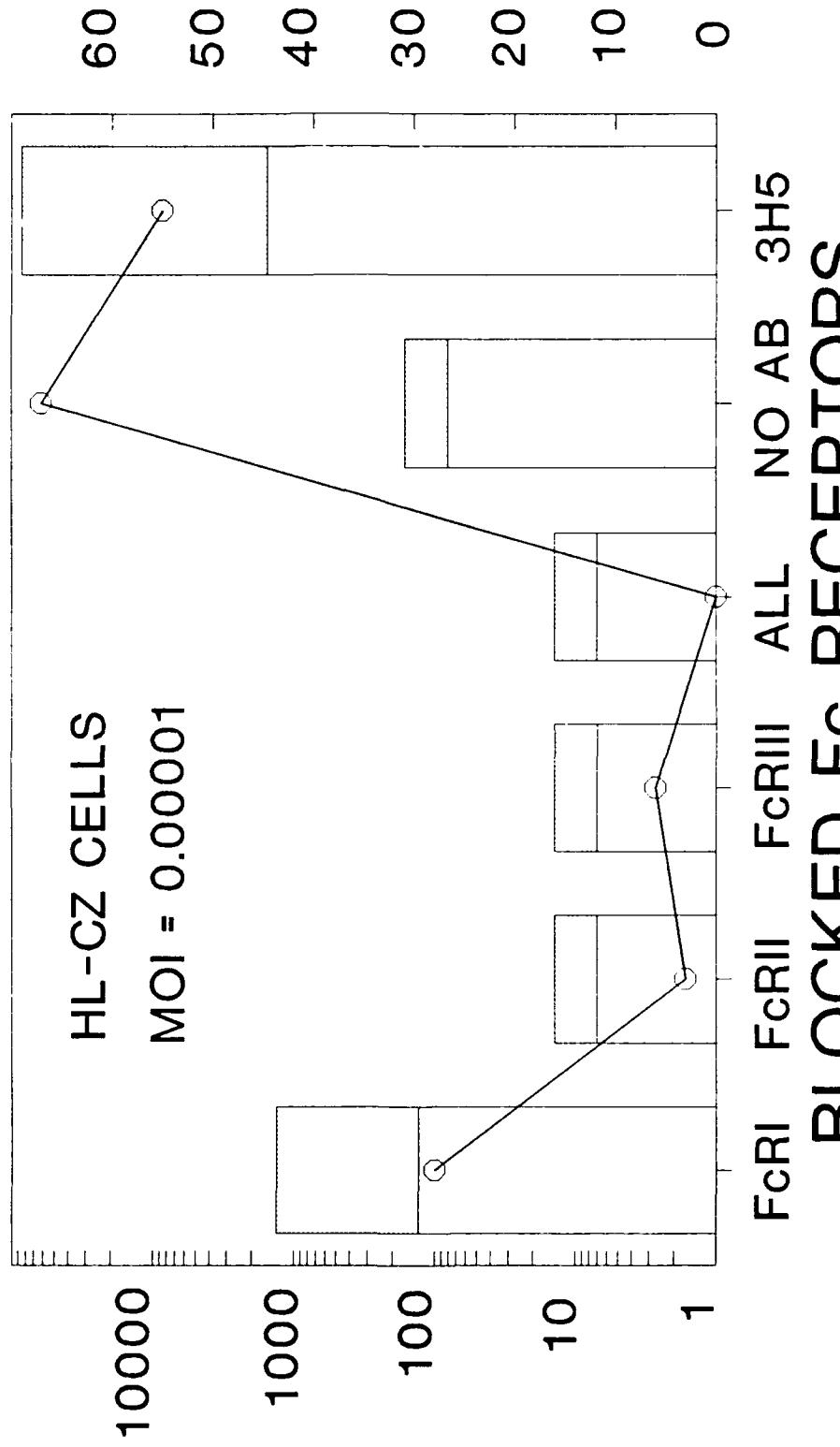


FIGURE 3. DAY 2 Harvest (open area), DAY 4 Harvest (shaded area)

SPECIFICITY OF ANTIBODY MEDIATING
ENHANCED DEN-2 REPLICATION

PFU/ML

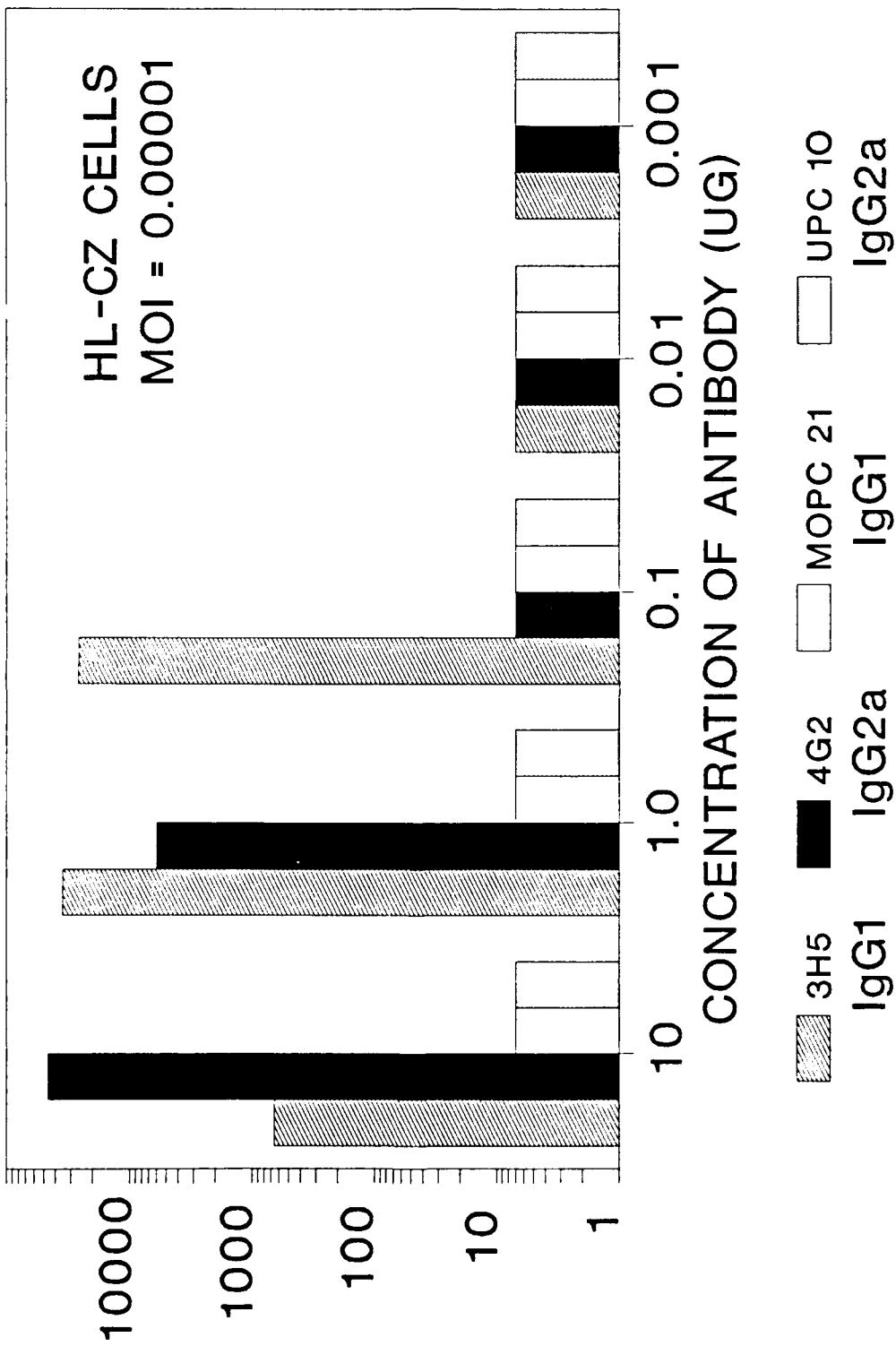
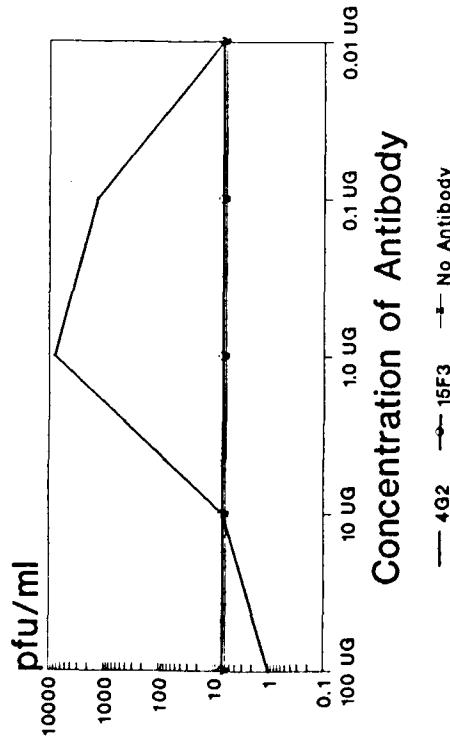


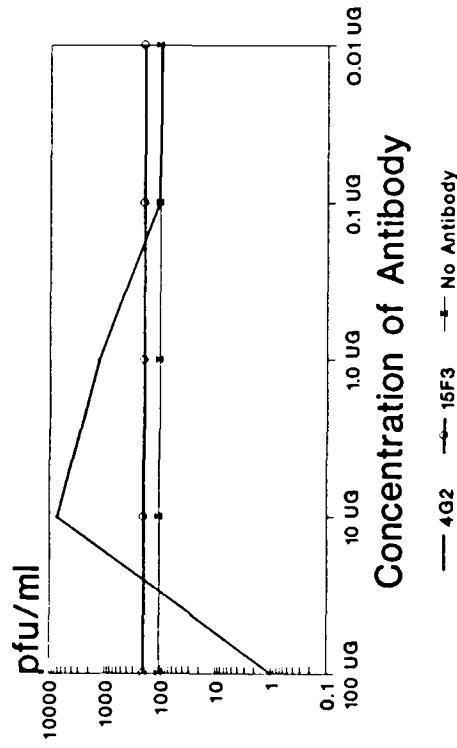
FIGURE 4.

THAILAND DEN-2 STRAIN 16681

THAILAND DEN-1 STRAIN 16007



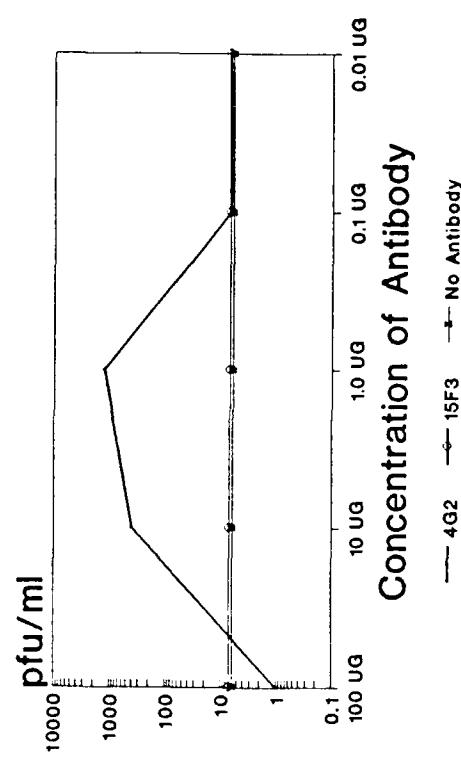
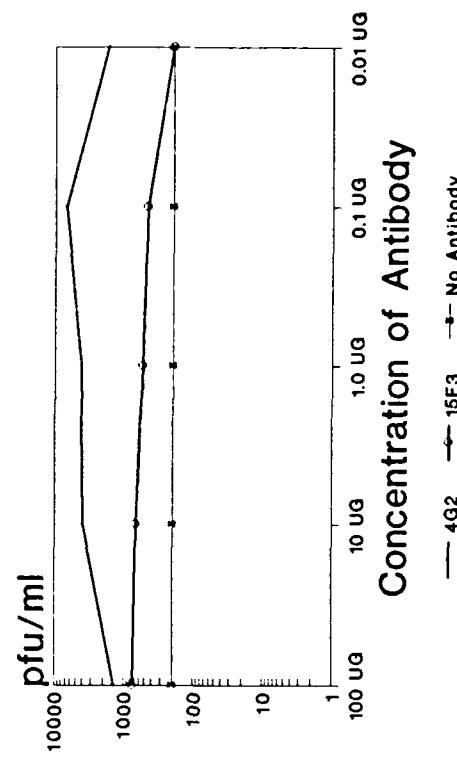
JAMAICA DEN-2 STRAIN 1409



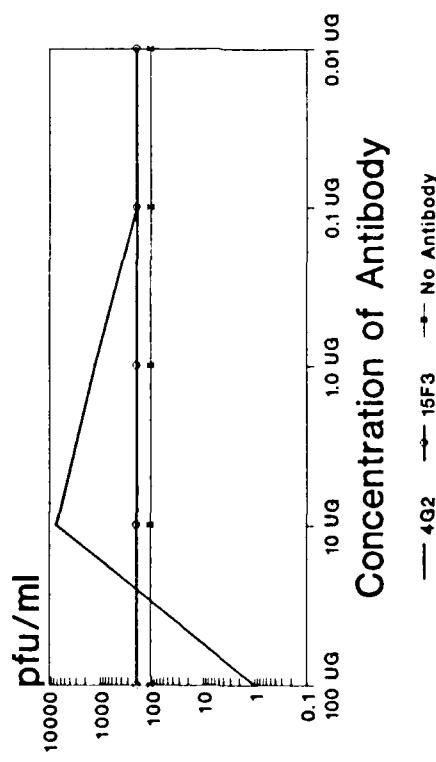
MOI = 0.0001

FIGURE 5.

JAMAICA DEN-1 STRAIN CV1636/77



JAMAICA DEN-2 STRAIN 1409
MOI = 0.0001



MOI = 0.00001

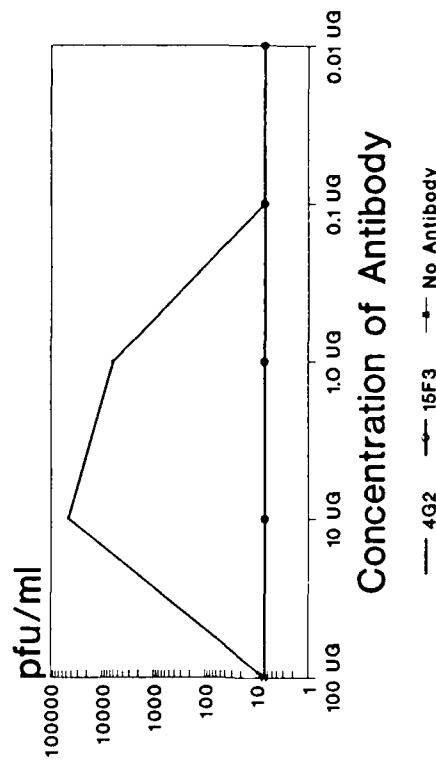


FIGURE 6.

ORGANIZATION OF THE DENGUE VIRUS GENOME

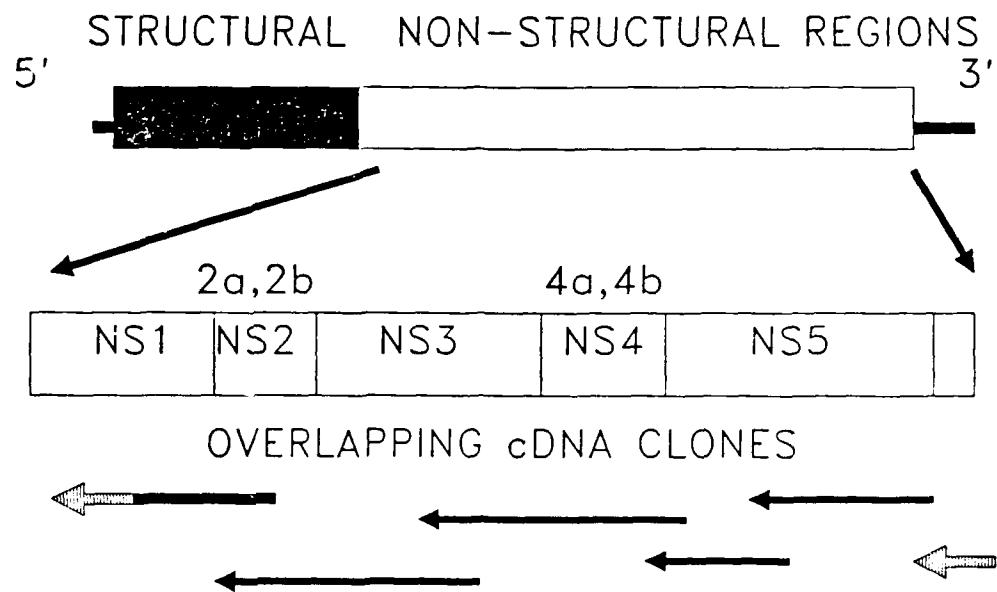


FIGURE 7.

DEDUCED AMINO ACID SEQUENCE CHANGES BETWEEN DENGUE VIRUSES

AA 5' NS4a GENOME 150 AMINO ACIDS 3'

DEN=2

DEM 4 11 1

5' NS4b GENOME 248 AMINO ACIDS 3

DEN-1

DEN-2

DEN-3

DEN-4

▲ AT POSITIONS 21-23
THERE ARE 3 AA
DELETIONS IN DEN-1, DEN-4

5' NSS GENOME 846/ 900 AA 3'
▲ DEF-1

DEN-2 ||| |||| ||| ||| ||| ||| |||

DEN-3 | | | | | | | | | | | | | | | |

▲ = BOTH DEN-1 AND DEN-3 HAVE A DELETION AT POSITION 176

FIGURE 8.